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Synthesis of 2-O-(3-O-Carbamoyl-α-D-mannopyranosyl)-L-gulopyranose: Sugar Moiety of Antitumor Antibiotic Bleomycin

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Abstract: A new route to the disaccharide moiety (2-O-(3-O-carbamoyl-α-D-mannopyranosyl)-L-gulopyranose) of the antitumor agent bleomycin was developed. Both the L-gulose synthon 21 and the 3-O-carbamoyl-D-mannose segment 30 were prepared from D-mannose in a regioselective manner by applying stannylene acetal methodology. Glycosylation of 21 with 30 proceeded smoothly, and further conversion to disaccharide derivatives (33 and 34) was successfully accomplished. © 1997 Elsevier Science Ltd.

The bleomycins (BLMs) are a family of glycopeptide antitumor antibiotics isolated from the fermentation broth of *Streptomyces verticillus* in 1966 by Umezawa *et al.*,¹ and currently used for the clinical treatment of Hodgkin's lymphoma, carcinomas of the skin, head, and neck, and tumors of the testis.² Cleavage of DNA by BLM in the presence of oxygen and ferrous ion occurs preferentially at G-C (5'→3') and G-T (5'→3') sequences, and is responsible for its antitumor activity.³ BLM is a glycopeptide having a unique hexapeptide and a disaccharide.⁴ We have been carrying out a synthetic approach to define the role of each segment through the synthesis of model compounds.⁵ Our attention has been focused on (1) the metal binding site, pyrimidoblamic acid connected to a hydroxyhistidine moiety,⁶ and (2) the DNA binding site, the bithiazole and linker moieties.³ Based on these studies of structure-function relationships, G-specific and AT-specific BLM analogues (PYML-6 and PYML(6)-(AHM)-distamycin, respectively) were successfully synthesized.³ Although these analogues showed potent DNA cleaving activity with the controlled sequence specificity, they were less cytotoxic against tumor cells in suspension culture than BLM. The significant decrease in cytotoxicity of BLM analogues in a cell system is apparently due to the lack of a disaccharide moiety in those analogues, in which a *tert*-butyl group is introduced instead. Because BLM localizes specifically in tumor cells in man and other animals³c.8, allowing 57Co-BLM to be used clinically as a cancer diagnostic drug,8b the

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disaccharide moiety of BLM has been hypothesized to be significant not only for the formation of a molecular cavity to accommodate dioxygen but also for molecular recognition at cell surfaces. All of the model compounds ("designed Bleomycins") we have prepared so far have a *tert*-butyl group, and by these models we could demonstrate the role of the disaccharide moiety as a bulky steric factor to accommodate dioxygen. However, this seems to be only one of several functions of the disaccharide moiety. In order to investigate another important role of the disaccharide moiety, *i.e.*, cell permeability and molecular recognition at cell surfaces, it became necessary to develop a practical method for the synthesis of the disaccharide, 2-O-(3-O-carbamoyl-α-D-mannopyranosyl)-L-gulopyranose.

In previous communications we reported (1) a novel approach to L-gulose from D-mannose, ¹⁰ (2) a regioselective preparation of 3-O-carbamoyl-D-mannose, ¹¹ and (3) an efficient coupling of each fragment and further transformation to the disaccharide moiety. ¹¹ This paper describes in full detail the synthesis of 2-O-(3-O-carbamoyl-α-D-mannopyranosyl)-L-gulopyranose.

Synthetic Strategy

Synthesis of the disaccharide moiety has been accomplished by three groups as a part of the total synthesis of BLM. In early work by Umezawa et al.¹² and Hecht et al.,¹³ the L-gulose skeleton was constructed from D-glucose based on the consideration that the interconversion of the oxidation level of C1 and C6 of D-glucose would give L-gulose. Alternatively, we envisioned that inversion of the stereochemistry at C5 of D-mannose would also provide a novel approach to L-gulose. Indeed, independent of our work, Boger et al.¹⁴ recently developed a stereoselective transformation of D-mannose to the L-gulose skeleton using a Rh(I)-catalyzed hydroboration of a 6-deoxy-hex-5-enopyranoside derivative during the course of their total synthesis of BLM,¹⁴,¹⁵

Our basic strategy is shown in Scheme 1. The key feature is that a linear allylic alcohol 1 was envisaged as an equivalent of L-gulose and was expected to be a potential glycosyl acceptor because the hydroxyl at C2¹⁶ is a reactive allylic hydroxyl.¹⁷ Heptose 1, in turn, might be obtained from a suitably protected D-mannopyranose 4 by Wittig reaction, followed by inversion of stereochemistry at C5 of the resulting 3. After glycosylation of 1 with mannose segment 2, ozonolysis of the exomethylene part would afford the disaccharide moiety of BLM.

Scheme 1

Disaccharide Moiety of Bleomycin

$$R^{1}O \xrightarrow{QR^{1}} QR^{2} \longrightarrow R^{1}O \longrightarrow$$

Transformation of D-Mannose to L-Gulose

Transformation of D-mannose to L-gulose was first examined with 2,3,4,6-tetra-O-benzyl-D-mannose 4a as a model. Treatment of 4a in THF with 2 equivalents of ylide generated from phosphonium bromide and n-BuLi gave the heptenol 3a in 65% yield along with the undesired dienol 5 in 8% yield (Scheme 2).

Scheme 2

In the case of the related D-glucose derivative, Sinaÿ et al.¹⁸ reported that the formation of enol and/or dienol depends on the solvent and base employed. Indeed, when the ylide was generated using potassium hexamethyldisilazide (KHMDS) as a base, dienol 5 was obtained exclusively in 80% yield. Further, the yield of 3a was improved to 87% by the pretreatment of 4a with an equimolar amount of n-BuLi followed by the reaction with Ph₃P=CH₂ (base: n-BuLi).

With heptose 3a in hand, S_N 2 type inversion at C5¹⁶ was initially attempted. Various methods, such as the Mitsunobu reaction¹⁹ or the substitution of sulfonates with CsOAc²⁰ or KO₂,²¹ gave the inversion product only in low yields (Scheme 3).

Scheme 3

We then became interested in an alternative oxidation-reduction approach *via* a 5-keto derivative. ¹⁶ Protection of O-4 and O-6 as a benzylidene acetal was considered most suitable for achieving high stereoselectivity because the 5-keto group becomes part of a 6-membered dioxanone ring. In a model study, both hydroxyl groups in benzylidene thiomannoside 7, prepared from 6 in 53% yield, were benzylated to provide fully protected thioglycoside 8 in 98% yield. Selective protection of O-2 or O-3 in 7 was also possible *via* its stannylene acetal, and that methodology was successfully utilized in the preparation of both the L-gulose and 3-*O*-carbamoyl-D-mannose segments (*vide infra*). Thioglycoside 8 was hydrolyzed with NBS in acetone-H₂O to afford hemiacetal 9, and Wittig olefination of the latter proceeded smoothly giving the 5α-hydroxy-1,3-dioxane 10²² in 65% yield (87% conversion yield based on recovered 9). Swern oxidation of dioxanol 10 gave the dioxanone 11 in 84% yield (Scheme 4).

Scheme 4

Dioxanone 11 was then subjected to reduction with various hydride reagents. As shown in Scheme 5, the desired β-alcohol 12 was exclusively formed when LiBH(sec-Bu)3 (L-Selectride) was used. Brown et al. reported that the equatorial attack by L-Selectride was particularly significant in the case of α-substituted cyclohexanones.²³ The small coupling constants of H-5 ($J_{5,6eq}$ =-0Hz, $J_{5,6ax}$ =2.0Hz and $J_{4,5}$ =1.1Hz) well supported the axial orientation of the hydroxyl group in 12. On the other hand, preferential axial attack was observed with NaBH4 and DIBAL. Conversion of the heptose 12 to L-gulose was accomplished in 46% overall yield by ozonolysis of the terminal olefin and hydrogenolysis of the benzylidene and benzyl groups (Scheme 6). In this way we have established a novel method for the conversion of D-mannose to L-gulose by inversion at C-5 through an oxidation-reduction sequence. 10

Scheme 5

90

>99

Preparation of L-Gulose Synthon

For the attachment of the D-mannose portion it was necessary to differentiate the gulose C2 hydroxyl. CsF-mediated regioselective alkylation of stannylene acetal 14^{24} was examined for this purpose. Boger et al. ¹⁴ had also utilized stannylene acetal methodology for the selective O-3 alkylation of the related mannose derivatives. Stannylene acetal 14, prepared from diol 7, was found to undergo regioselective benzylation at room temperature in the presence of CsF to afford O-3 benzylation product 15 as a single isomer. The structure of 15 was confirmed after acetylation to 16. In the ¹H-NMR spectrum of acetate 16, H-2 was shifted downfield; δ 4.29 (H-2; $J_{1,2}$ =1.0, $J_{2,3}$ =3.4 Hz) and δ 3.97 (H-3; $J_{2,3}$ =3.4, $J_{3,4}$ =9.5Hz) for 15; δ 5.62 (H-2; $J_{1,2}$ =1.4, $J_{2,3}$ =3.4 Hz) and δ 3.86 (H-3; $J_{2,3}$ =3.4, $J_{3,4}$ =9.8 Hz) for 16.

Scheme 7

Scheme 8

The remaining hydroxyl at C2 of 15 was protected as an MPM (4-methoxybenzyl) ether, and thioglycoside 17 was hydrolyzed to mannopyranose 18 in 54% yield (91% conversion yield based on recovered 17). Methylenation of 18 was carried out using Ph₃P=CH₂ to obtain heptose 19 in 66% yield. Following the protocol established in the model study described above, D-manno-type 19 was converted to L-gulo-type 20 in 83% overall yield with complete stereoselectivity. After acetylation of dioxanol 20, the MPM group was selectively deprotected²⁵ to obtain allylic alcohol 21 which serves as a glycosyl acceptor (Scheme 7). When dioxanol 20 was treated with DDQ, the desired diol 22 was obtained only in low yield, and the acetal 23 was isolated in high yield instead (Scheme 8). Therefore, the dioxanol hydroxyl was acetylated prior to the deprotection of the MPM group. Related oxidative acetalization was also reported by Yonemitsu *et al.*²⁶

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Preparation of 3-O-Carbamoyl-D-Mannose Portion

In the preparation of L-gulose synthon we described the regioselective benzylation at C3-OH of 4,6-O-benzylidene mannose 7 through its stannylene acetal. The same strategy was also applied for the preparation of the 3-O-carbamoyl-D-mannose moiety (Scheme 9). In this case 4-methoxybenzyl group (MPM) was introduced instead at O-3. Thus, the stannylene acetal 14 derived from 7 was reacted with MPMBr, prepared in situ from MPMCl and n-Bu₄NBr, in the presence of CsF to afford 24 in 94% yield as a single isomer. The regiochemistry of 24 was tentatively assigned in analogy to that of 15, and was confirmed after transforming to the carbamoyl derivative 28. The benzylidene group was hydrolyzed, and the resulting triol 25 was benzylated to obtain 26 almost quantitatively. Then the MPM group was selectively deprotected with DDQ to generate the C3-OH. The introduction of a carbamoyl group was carried out in 92% yield by a conventional method, p-nitrophenyl carbonation followed by ammonolysis. The 3-O-carbamoyl structure was confirmed by NMR. Thus, in the 1 H-NMR spectrum of carbamoyl mannoside 28, H-3 and H-2 appeared at δ 5.13 (dd, $J_{2,3}$ =3.2, $J_{3,4}$ =9.2Hz) and δ 4.14 (dd, $J_{2,3}$ =3.2, $J_{1,2}$ =2.1Hz), respectively. Thioglycoside 28 was hydrolyzed with NBS, and the resulting 29 was reacted with Cl₃CCN and a catalytic amount of DBU to obtain 30 in 92% yield.²⁷ Trichloroacetimidate 30 serves as a glycosyl donor.

Scheme 9

Boger et al. also established an efficient route to 2,4,6-tri-O-acetyl-3-O-carbamoyl mannose (not shown) employing a similar stannylene acetal methodology. 14 Our approach, although it seems complementary to that of Boger et al., is quite useful because the disaccharide moiety can be prepared in a differently protected manner.

We were thus able to prepare both L-gulose and D-mannose segments from the common intermediate 7, and the stannylene acetal methodology was successfully applied to achieve the regioselective alkylation of 4,6-O-benzylidene mannoside 7.

Coupling of Two Segments and Synthesis of Disaccharide

With both glycosyl acceptor and donor in hand, the coupling of 21 and 30 was then examined. When allyl alcohol 21 was treated with trichloroacetimidate 30 in CH₂Cl₂ in the presence of BF₃•OEt₂, glycosylation

proceeded smoothly at -15°C within 10 min to isolate the desired α -mannosyl derivative 31 in 82% yield (Scheme 10). The high reactivity of 21 is noteworthy and is apparently derived by masking the C1 carbonyl as a vinyl group.

Scheme 10

After deacetylation, the resulting hydroxy olefin 32 was subjected to ozonolysis to obtain 33 as an anomeric mixture (α : β =ca 2:1). Hydrogenolysis using Pd(OH)₂ gave free 2-(3-O-carbamoyl- α -D-mannopyranosyl)-L-gulose, which was converted to the heptaacetate 34 in 89% yield as a single β -anomer ($J_{1,2}$ =8.4 Hz). Spectral data (1 H-and 13 C-NMR) of 34 were identical to those kindly provided by Prof. D. L. Boger (The Scripps Research Institute). Selective deprotection of the anomeric acetate in 34 has already been established by Boger et al. 14

In conclusion, we have developed a new route to the disaccharide moiety of BLM. The key features of the present synthesis are as follows: (1) the L-gulose skeleton is stereoselectively constructed from D-mannose by inversion of stereochemistry at C5 through an oxidation-reduction sequence; (2) methylenation of the C1 carbonyl group of 4 not only served as temporary protection but also resulted in increased nucleophilicity of the C2 hydroxyl group; (3) stannylene acetal methodology was effectively applied for the preparation of both fragments. Further, it should also be emphasized that both 33 and 34 are considered versatile intermediates for introducing the disaccharide moiety into diverse types of compounds. Protecting groups in 33 (Bn) and 34 (Ac) may be removed under quite different conditions after introduction of a given aglycon. Since the disaccharide moiety of BLM is hypothesized to play a central role in molecular recognition at cell surfaces and also in cell permeability, the biological properties of such hybrid compounds bearing the disaccharide are likely to prove quite interesting. Synthetic efforts along these lines are now in progress.

Experimental Section

All melting points were determined with a Yanagimoto MP-21 melting point apparatus and were uncorrected. Optical rotations were measured with a Horiba SEPA-200 auto digital polarimeter. Infrared (IR) spectra were recorded on a JASCO A-202 spectrometer. 1 H-NMR spectra were measured with a Bruker AC 200P (200MHz), and a Bruker AM 400 (400MHz) spectrometer. The chemical shifts are expressed in ppm downfield from tetramethylsilane, using tetramethylsilane (δ =0) and/or residual chloroform (δ =7.25) as an internal standard. Splitting patterns are indicated as s, singlet; d, doublet; t, triplet; q, quartet; m, multiplet; br, broad signal. Elemental analysis was performed with Perkin-Elmer 240 or 2400. Mass spectra were taken with a Hitachi M-80A or M-80B mass spectrometer. Unless otherwise noted, all experiments were carried out under an atmosphere of dry argon using anhydrous solvents. For thin layer chromatographic (TLC) analyses, Merck precoated TLC plates (silica gel 60 F254, 0.25mm, Art 5715) were used. The following abbreviations were used for solvents: tetrahydrofuran (THF), diethyl ether (Et₂O), ethyl acetate (AcOEt), methanol (MeOH), and dichloromethane (CH₂Cl₂).

(2R,3R,4R,5R)-1,3,4,5-Tetrabenzyloxy-6-hepten-2-ol (3a): To a solution of 4a (1.215 g, 2.25 mmol) in THF (10 mL) was added a 1.6 *M* hexane solution of n-BuLi (1.45 mL, 2.33 mmol) at -78°C. The mixture was stirred at 0°C for 5 min and then cooled to -78°C. To the above solution was added the THF solution (10 mL) of Ph₃P=CH₂, prepared from Ph₃PCH₃+Br⁻ (0.889 g, 2.49 mmol) and n-BuLi (1.6*M* hexane solution, 1.70 mL, 2.73 mmol), and the mixture was stirred at r.t. for 30 hr. The reaction mixture was poured into sat. NaHCO₃, and the products were extracted with AcOEt. The organic layer was washed with brine, dried over anhydrous Na₂SO₄, and concentrated under reduced pressure. The residue was chromatographed on silica gel (AcOEt/hexane=1/9) to give 3a (1.06 g, 87%) as a colorless oil. $[\alpha]_D^{20}$ +7.2° (*c* 1.00, CHCl₃): IR (neat) 3030, 2912, 2866, 1496, 1454cm⁻¹: ¹H-NMR (400MHz, CDCl₃) δ 2.64 (1H, d, *J*=4.8Hz, OH), 3.63 (1H, dd, *J*=3.2, 9.8Hz), 3.66 (1H, dd, *J*=5.5, 9.8Hz), 3.84-3.89 (2H, m), 4.00 (1H, m), 4.11 (1H, m), 4.24 (1H, d, *J*=11.5Hz), 4.44-4.65 (7H, m), 4.71 (1H, d, *J*=11.0Hz), 5.40 (1H, dd, *J*=1.5, 10.1Hz), 5.44 (1H, dd, *J*=1.1, 17.4Hz), 5.96 (1H, ddd, *J*=8.0, 10.1, 17.4Hz), 7.20-7.30 (20H, m): EI-MS m/z 539 ([M+1]+).

Phenyl 4,6-O-benzylidene-1-thio- α -D-mannopyranoside (7): 6 (6.41 g, 14.6 mmol) was dissolved in MeOH (70 mL) saturated with NH₃. The mixture was stirred at r.t. overnight, and the removal of the solvent gave the crude phenyl 1-thio- α -D-mannopyranoside. The crude material was dissolved in DMF (20 mL), and to the mixture was added successively benzaldehyde dimethylacetal (2.18 mL, 14.5 mmol) and HBF₄•OEt₂ (85% Et₂O solution, 1.68 mL, 11.4 mmol) at 0°C. After being stirred at r.t. overnight, Et₃N (2.5 mL) was added, and the mixture was poured into H₂O (150 mL). The precipitate was filtered, washed with cold Et₂O, and dried under vacuum to give 7 (2.79 g, 53%) as white crystals. m.p. 213-214°C (AcOEt): $[\alpha]_{2}^{2}$ 0 +289° (c 0.50, CHCl₃/MeOH=1/1): IR (KBr) 3346, 1452cm⁻¹: ¹H-NMR (200MHz, CDCl₃/CD₃OD=10/1) δ 3.85 (1H, t, J=10.1Hz), 3.96-4.12 (2H, m), 4.17-4.40 (3H, m), 5.57 (1H, d, J=0.9 Hz,), 5.59 (1H, s), 7.25-7.55 (10H, m).

Phenyl 2,3-di-O-benzyl-4,6-O-benzylidene-1-thio-α-D-mannopyranoside (8): DMF solution (50 mL) of 7 (8.00 g, 22.2 mol) was added to a suspension of NaH (ca 60% in mineral oil, 2.36 g, 59.1 mmol) in DMF (30 mL) at 0°C, and the mixture was stirred for 30 min. To the mixture was added benzyl bromide (7.14 mL, 59.9 mmol), and the reaction mixture was stirred at r.t. for 8 hr. MeOH (6 mL) was added and the mixture was poured into H₂O. The products were extracted with Et₂O, and the organic layer was chromatographed on silica gel (AcOEt/hexane=1/9) to give 8 (11.8 g, 98%) as white crystals. m.p. 84-85°C (AcOEt/Et₂O): [α]²⁰_D +107° (c 1.13, CHCl₃): IR (KBr) 3034, 2895, 2864, 1581, 1454cm⁻¹: ¹H-NMR (400MHz, CDCl₃) δ 3.89 (1H, t, J=9.5Hz), 3.97 (1H, dd, J=3.2, 9.5Hz), 4.04 (1H, dd, J=1.4, 3.2Hz), 4.20-4.35 (3H, m), 4.66 (1H, d, J=12.2Hz), 4.73 (2H, br.s), 4.83 (1H, d, J=12.2Hz), 5.51 (1H, d, J=1.4Hz), 5.65 (1H, s), 7.25-7.41 (18H, m), 7.52 (2H, m): Anal. Calcd for C₃₃H₃₂O₅S: C, 73.31; H, 5.97; S, 5.93%. Found: C, 73.33; H, 6.06; S, 5.92%.

2,3-Di-O-benzyl-4,6-O-benzylidene-D-mannopyranose (9): **8** (11.3 g, 20.9 mmol) was dissolved in acetone- H_2O (24/1, 80 mL), and was added dropwise a acetone- H_2O (24/1, 80 mL) solution of NBS (22.2 g, 125 mmol) at 0°C. The mixture was stirred at r.t. for 25 min, and the reaction was quenched with sat. Na₂SO₃. The products were extracted with Et₂O, and the organic layer was successively washed with H_2O and brine, dried over anhydrous MgSO₄, and concentrated under reduced pressure. The residue was chromatographed on silica gel (AcOEt/hexane) to give **9** (5.80 g, 62%) as an anomeric mixture. ¹H-NMR (400MHz, CDCl₃) δ 2.65 (1H, d, J=3.4 Hz, OH), 3.86 (1H, dd, J=1.6, 3.1Hz), 3.86 (1H, t, J=10.1Hz), 4.02 (1H, dd, J=3.1, 9.8Hz), 4.02 (1H, ddd, J=4.6, 9.8, 10.1Hz), 4.23 (1H, dd, J=4.6, 10.1Hz), 4.26 (1H, t, J=9.8Hz), 4.67 (1H, d, J=12.2Hz), 4.73 (1H, d, J=12.2Hz), 4.83 (1H, d, J=12.2Hz), 4.84 (1H, d, J=12.2Hz), 5.19 (1H, dd, J=1.6, 3.4Hz), 5.64 (1H, s), 7.26-7.41 (18H, m), 7.50 (2H, m): *Anal.* Calcd for C₂₇H₂₈O₆: C, 72.30; H, 6.29%. Found: C, 72.10; H, 6.51%.

(1'R,2R,2'R,4R,5R)-4-(1,2-Dibenzyloxy-3-butenyl)-2-phenyl-1,3-dioxan-5-ol (10): 9 (1.63 g, 3.62 mmol) was reacted with Ph₃P=CH₂ in a manner similar to the preparation of **3a** to give **10** (1.06 g, 65%, and 87% conversion yield based on recovered **9** (0.40 g)) as white crystals. m.p. 85°C (Et₂O/hexane): $[\alpha]_0^{20}$ -48.2° (c 1.04, CHCl₃): IR (KBr) 3476, 2903, 2870, 1643, 1452cm⁻¹: ¹H-NMR (400MHz, CDCl₃) 8 1.70 (1H, d, J=4.0Hz, OH), 3.52 (1H, dd, J=9.3, 9.4Hz), 3.84-3.94 (3H, m), 4.22 (1H, dd, J=4.8, 10.4Hz), 4.23 (1H, dd, J=6.8, 7.4Hz), 4.38 (1H, d, J=11.7Hz), 4.59 (1H, d, J=11.7Hz), 4.66 (1H, d, J=11.7Hz), 4.79 (1H, d, J=11.7Hz), 5.37 (1H, s), 5.44 (1H, ddd, J=0.6, 1.6, 10.3Hz), 5.47 (1H, ddd, J=0.7, 1.6, 17.7Hz), 6.05 (1H, ddd, J=7.6, 10.3, 17.7Hz), 7.23-7.40 (15H, m): *Anal*. Calcd for C₂₈H₃₀O₅: C, 75.31; H, 6.77%. Found: C, 75.40; H, 6.82%.

(1'R, 2R, 2'R, 4R)-4-(1,2-Dibenzyloxy-3-butenyl)-2-phenyl-1,3-dioxan-5-one (11): DMSO (0.28 mL, 3.95 mmol) was added to a CH₂Cl₂ solution (1.5 mL) of oxalyl chloride (0.17 mL, 1.95 mmol) at -50°C, and the mixture was stirred at that temperature for 15 min. Then 10 (369.7 mg, 0.83 mmol) in CH₂Cl₂ (1.5 mL) was added at -50°C. After being stirred at -50°C for 30 min and at -18°C for 10 min, Et₃N (1.20 mL, 8.61 mmol) was added at -18°C. After being stirred at -18°C for 10 min and at 0°C for 10 min, the reaction mixture was poured into sat. NaHCO₃. The products were extracted with Et₂O, and the organic layer was washed with brine, dried over anhydrous Na₂SO₄, and concentrated under reduced pressure. The residue was chromatographed on silica gel (AcOEt/hexane) to give 11 (309.2 mg, 84%) as a syrup. ¹H-NMR (400MHz, CDCl₃) δ 4.13 (1H, dd, J=2.0, 8.9Hz), 4.18 (1H, dd, J=7.2, 8.9Hz), 4.35 (1H, d, J=11.7Hz), 4.42 (2H, d, J=11.0Hz), 4.51 (1H, d, J=11.0Hz), 4.62 (1H, d, J=11.0Hz), 4.63 (1H, d, J=11.7Hz), 4.91 (1H, d, J=2.0Hz), 5.41 (1H, d, J=10.3Hz), 5.46 (1H, d, J=17.4Hz), 5.85 (1H, s), 5.91 (1H, ddd, J=7.2, 10.3, 17.4Hz), 7.20-7.49 (15H, m).

(1'R,2R,2'R,4R,5S)-4-(1,2-Dibenzyloxy-3-butenyl)-2-phenyl-1,3-dioxan-5-ol (12): A THF solution of L-Selectride (1.0 M, 1.0 mL, 1.0 mmol) was added to a THF solution (4 mL) of 11 (309.2 mg, 0.69 mmol) at -78°C. The mixture was stirred at -78°C for 90 min, then at 0°C for 30 min. The reaction was quenched with brine, and the products were extracted with AcOEt. The organic layer was washed with brine, dried over anhydrous Na₂SO₄, and concentrated under reduced pressure. The residue was chromatographed on silica gel (AcOEt/hexane) to give 12 (279.5 mg, 90%) as white crystals. m.p. 101-102°C (Et₂O/hexane): $[\alpha]_D^{20}$ -30.6° (c 0.98, CHCl₃): IR (KBr) 3485, 2914, 2872, 1496, 1454cm⁻¹: ¹H-NMR (400MHz, CDCl₃) δ 3.13 (1H, d, J=9.6Hz, OH), 3.65 (1H, dddd, J=1.1, 1.4, 2.0, 9.6Hz), 3.98 (1H, dd, J=4.2, 7.3Hz), 4.00 (1H, dd, J=1.4, 11.9Hz), 4.03 (1H, dd, J=1.1, 7.3Hz), 4.12 (1H, dd, J=4.2, 8.1Hz), 4.18 (1H, dd, J=2.0, 11.9Hz), 4.43 (1H, d, J=11.8Hz), 4.66 (1H, d, J=11.8Hz), 4.75 (1H, d, J=11.0Hz), 4.80 (1H, d, J=11.0Hz), 5.35 (1H, ddd, J=0.9, 1.7, 17.3Hz), 5.40 (1H, ddd, J=0.6, 1.7, 10.3Hz), 5.57 (1H, s), 6.04 (1H, ddd, J=8.1, 10.3, 17.3Hz), 7.23-7.40 (13H, m), 7.50 (2H, m): Anal. Calcd for C₂₈H₃₀O₅: C, 75.31; H, 6.77%. Found: C, 75.33; H, 6.75%.

Transformation of 12 to L-Gulose: Ozone gas was passed into a MeOH solution (10 mL) of 12 (194.5 mg, 0.44 mmol) at -78°C for 40 min until the solution became blue. Ar gas was then passed into the solution to remove an excess ozone. Me₂S (0.30 mL) was added to the reaction mixture, and the mixture was allowed to warm to r.t.. The mixture was concentrated under reduced pressure, and the residue was

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chromatographed on silica gel (AcOEt/hexane) to give 13 (168.5 mg, 86%) as an anomeric mixture. A mixture of 13 (15.2 mg, 0.03 mmol) and 20% Pd(OH)₂ (15 mg) in MeOH (10 mL) was stirred under H₂ (20 atm) in a sealed tube for 3 hr. The catalyst was filtered off, and the removal of the solvent gave L-gulose (3.1 mg, 56%) as a solid. 1 H-NMR (400MHz, CD₃OD) δ 3.56 (1H, dd, J=3.3, 8.2Hz), 3.68 (1H, dd, J=5.5, 11.3Hz), 3.71 (1H, m), 3.73 (1H, dd, J=6.5, 11.3Hz), 3.92 (1H, ddd, J=1.0, 5.5, 6.5Hz), 3.95 (1H, t, J=3.3Hz), 4.80 (1H, d, J=8.2Hz).

Phenyl 3-O-benzyl-4,6-O-benzylidene-1-thio-α-D-mannopyranoside (15): A mixture of 7 (0.859 g, 2.38 mmol) and Bu₂SnO (0.605 g, 2.43 mmol) in toluene (30 mL) was heated under refluxing for 3 hr. The solvent was removed under reduced pressure to obtain crude 14. The mixture of crude 14, CsF (0.369 g, 2.43 mmol) and benzyl bromide (0.29 mL, 2.44 mmol) in DMF (10 mL) was stirred at r.t. for 12 hr. The reaction mixture was poured into sat. NaHCO₃, and the products were extracted with Et₂O. The organic layer was washed with brine, dried over anhydrous Na₂SO₄, and concentrated under reduced pressure. The residue was chromatographed on silica gel (AcOEt/hexane) to give 15 (0.981 g, 91%) as white crystals. m.p. 89-90°C (Et₂O/hexane): [α]_D²⁰ +228° (c 1.07, CHCl₃): IR (KBr) 3454, 2899, 1736cm⁻¹: ¹H-NMR (400MHz, CDCl₃) δ 2.83 (1H, d, J=1.3Hz, OH), 3.86 (1H, t, J=10.3Hz), 3.97 (H, dd, J=3.4, 9.5Hz), 4.18 (1H, t, J=9.5Hz), 4.21 (1H, dd, J=4.9, 10.3Hz), 4.29 (1H, ddd, J=1.0, 1.3, 3.4Hz), 4.34 (1H, ddd, J=4.9, 9.5, 10.3Hz), 4.75 (1H, d, J=11.8Hz), 4.90 (1H, d, J=11.8Hz), 5.60 (1H, d, J=1.0Hz), 5.63 (1H, s), 7.28-7.53 (15H, m): Anal. Calcd for C₂₆H₂₆O₅S: C, 69.31; H, 5.82; S, 7.12%. Found: C, 69.07; H, 5.90; S, 6.83%.

Phenyl 3-O-benzyl-4,6-O-benzylidene-2-O-(4-methoxybenzyl)-1-thio-α-D-mannopyranoside (17): A DMF solution (9 mL) of 15 (0.853 g, 1.89 mmol) was added to a suspension of NaH (ca 60% in mineral oil, 83.5 mg, 2.09 mmol) in DMF (1 mL) at 0°C. After being stirred at 0°C, the mixture was added MPMCl (0.29 mL, 2.06 mmol), and was stirred at r.t. for 12 hr. MeOH and H₂O (30 mL) was added, and the products were extracted with Et₂O. The organic layer was washed with brine, dried over anhydrous MgSO₄, and concentrated under reduced pressure. The residue was chromatographed on silica gel (AcOEt/hexane) to give 17 (1.019 g, 94%) as a syrup. $[\alpha]_D^{20}$ +112° (c 0.66, CHCl₃): IR (neat) 3063, 2901, 2864, 1612cm⁻¹: ¹H-NMR (400MHz, CDCl₃) δ 3.79 (3H, s), 3.88 (1H, dd, J=9.5, 9.5Hz), 3.95 (1H, dd, J=3.2, 9.5Hz), 4.02 (1H, dd, J=1.4, 3.2Hz), 4.19-4.32 (3H, m), 4.64 (1H, d, J=12.1Hz), 4.65 (2H, br.s), 4.81 (1H, d, J=12.1Hz), 5.46 (1H, d, J=1.4Hz), 5.64 (1H, s), 6.84 (2H, m), 7.25-7.40 (15H, m), 7.52 (2H, m): Anal. Calcd for C₃₄H₃₄O₆S: C, 71.56; H, 6.01; S, 5.62%. Found: C, 71.28; H, 6.05; S, 5.47%.

3-O-Benzyl-4,6-O-benzylidene-2-O-(4-methoxybenzyl)-D-mannopyranose (18): NBS (127.3 mg, 0.72 mmol) was added to 17 (388.6 mg, 0.68 mmol) in acetone(15 mL) containing H₂O (0.3 mL) at 0°C, and the mixture was stirred at r.t. for 30 min. The reaction was quenched with sat. Na₂SO₃, and the products were extracted with Et₂O. The organic layer was washed with brine, dried over anhydrous MgSO₄, and concentrated under reduced pressure. The residue was chromatographed on silica gel (AcOEt/hexane) to give 18 (175.0 mg, 54%, and 91% conversion yield based on recovered 17 (158.2 mg)) as an anomeric mixture. 1 H-NMR (400MHz, CDCl₃) δ 2.66 (1H, d, J=3.5Hz, OH), 3.78 (3H, s), 3.84 (1H, dd, J=1.6, 3.1Hz), 3.85 (1H, t, J=10.2Hz), 4.00 (1H, dd, J=3.1, 9.8Hz), 4.02 (1H, ddd, J=3.1, 4.8, 9.7Hz), 4.22 (1H, dd, J=4.8, 10.2Hz), 4.24 (1H, dd, J=9.7, 9.8Hz), 4.65 (1H, d, J=12.2Hz), 4.66 (1H, d, J=12.0Hz), 4.76 (1H, d, J=12.0Hz), 4.83 (1H, d, J=12.2Hz), 5.14 (1H, dd, J=1.6, 3.5Hz), 5.64 (1H, s), 6.85 (2H, m), 7.24-7.52 (12H, m).

(1'R,2R,2'R,4R,5R)-4-[(1-Benzyloxy-2-(4-methoxybenzyl)oxy-3-butenyl]-5-hydroxy-2-phenyl-1,3-dioxane (19): 18 (6.90 g, 14.4 mmol) was reacted with Ph₃P=CH₂ in a manner similar to the preparation of 3a to give 19 (4.56 g, 66%) as white crystals. m.p. 100-102°C (Et₂O/hexane): $[\alpha]_D^{20}$ -56.2° (c 1.35, CHCl₃): IR (KBr) 3437, 2932, 2866, 1612cm⁻¹: ¹H-NMR (400MHz, CDCl₃) δ 1.71 (1H, d, J=4.0 Hz, OH), 3.51 (1H, dd, J=9.5, 10.8Hz), 3.72 (3H, s), 3.80-3.90 (3H, m), 4.22 (1H, dd, J=4.8, 10.4Hz), 4.23 (1H, dd, J=6.8, 7.7Hz), 4.31 (1H, d, J=11.4Hz), 4.58 (1H, d, J=11.7Hz), 4.59 (1H, d, J=11.4Hz), 4.76 (1H, d, J=11.7Hz), 5.35 (1H, s), 5.44 (1H, dd, J=1.7, 9.8Hz), 5.47 (1H, dd, J=1.7, 17.4Hz), 6.03 (1H, ddd, J=7.6, 9.8, 17.4Hz), 6.79 (2H, m), 7.22 (2H, m), 7.31-7.39 (10H, m): Anal. Calcd for C₂₉H₃₂O₆: C, 73.09; H, 6.77%. Found: C, 73.02; H, 6.83%.

(1'R,2R,2'R,4R,5S)-4-[(1-Benzyloxy-2-(4-methoxybenzyl)oxy-3-butenyl]-5-hydroxy-2-phenyl-1,3-dioxane (20): 19 (2.33 g, 4.90 mmol) was oxidized with oxalyl chloride (0.94 mL, 11 mmol), DMSO (1.53 mL, 22 mmol) and Et₃N (3.41 mL, 24.5 mmol) in CH₂Cl₂ (55 mL) in a manner similar to the preparation of 11. Resulting crude dioxanone (2.32 g) was reduced with L-Selectride (1.0 M THF solution, 7.34 mL, 7.34 mmol) in a manner similar to the preparation of 12 to give 20 (1.94 g, 83% from 19) as white crystals. m.p. 109-110°C (Et₂O/hexane): $[\alpha]_D^{20}$ -33.8° (c 0.88, CHCl₃): IR (KBr) 3489, 2905, 2068, 1614, 1518, 1452cm⁻¹: ¹H-NMR (400MHz, CDCl₃) 8 3.17 (1H, d, J=9.6Hz, OH), 3.65 (1H, dddd, J=1.1, 1.4, 2.0, 9.6Hz), 3.79 (3H, s), 3.93 (1H, dd, J=4.6, 7.1Hz), 4.00 (1H, dd, J=1.4, 11.9 Hz), 4.03 (1H, dd, J=1.1, 7.1Hz), 4.10 (1H, dd, J=4.6, 8.0Hz), 4.19 (1H, dd, J=2.0, 11.9Hz), 4.36 (1H, dd, J=11.6Hz), 4.59 (1H, d, J=11.6Hz), 4.73 (1H, d, J=11.0Hz), 4.77 (1H, d, J=11.0Hz), 5.34 (1H, ddd, J=0.8, 1.7, 17.3Hz), 5.40 (1H, dd, J=0.6, 1.7, 10.3Hz), 5.57 (1H, s), 6.03 (1H, ddd, J=8.0, 10.3, 17.3 Hz), 6.85 (2H, m), 7.25-7.38 (10H, m), 7.56 (2H, m): Anal. Calcd for C₂9H₃₂O₆: C, 73.09; H, 6.77%. Found: C, 73.06: H, 6.88%.

(1'R,2R,2'R,4R,5S)-5-Acetoxy-4-[1-benzyloxy-2-(4-methoxybenzyl)oxy-3-butenyl]-2-phenyl-1,3-dioxane: A mixture of 20 (153.5 mg, 0.322 mmol), Ac₂O (0.5 mL) and pyridine (2 mL) was stirred at r.t. for 6 hr. The reaction mixture was diluted with Et₂O (30 mL), washed successively with sat. NaHCO₃ and brine, and dried over anhydrous Na₂SO₄. The Et₂O solution was concentrated under reduced pressure, and the residue was chromatographed on silica gel (AcOEt/hexane) to give (1'R,2R,2'R,4R,5S)-5-acetoxy-4-[1-benzyloxy-2-(4-methoxybenzyl)oxy-3-butenyl]-2-phenyl-1,3-dioxane (157.5 mg, 94%) as a colorless oil. [α]²⁰ -6.87° (c 1.25, CHCl₃): IR (neat) 2914, 2866, 1738, 1612, 1514, 1454cm⁻¹: ¹H-NMR (400MHz, CDCl₃) δ 1.99 (3H, s), 3.79 (1H, dd, J=3.1, 8.1Hz), 3.80 (3H, s), 3.95 (1H, dd, J=3.1, 8.4Hz), 4.00 (1H, dd, J=1.6, 13.0Hz), 4.07 (1H, dd, J=1.6, 8.4Hz), 4.28 (1H, dd, J=1.6, 13.0Hz), 4.31 (1H, d, J=11.8Hz), 4.57 (1H, d, J=11.8Hz), 4.72 (1H, q, J=1.6Hz), 4.79 (1H, d, J=11.3Hz), 4.82 (1H, d, J=11.3Hz), 5.30 (1H, ddd, J=0.8, 1.5, 17.4Hz), 5.37 (1H, ddd, J=0.6, 1.5, 10.3Hz), 5.59 (1H, s), 5.96 (1H, ddd, J=8.1, 10.4, 17.4Hz), 6.87 (2H, m), 7.21-7.40 (10H, m), 7.51 (2H, m): Anal. Calcd for C₃₁H₃₄O₇: C, 71.80; H, 6.61%. Found: C, 71.70; H, 6.78%.

(1'R,2R,2'R,4R,5S)-5-Acetoxy-4-(1-benzyloxy-2-hydroxy-3-butenyl)-2-phenyl-1,3-dioxane (21): To a solution of (1'R,2R,2'R,4S,5S)-5-acetoxy-4-[1-benzyloxy-2-(4-methoxybenzyl)oxy-3-butenyl]-2-phenyl-1,3-dioxane (593 mg, 1.14 mmol) in CH₂Cl₂-H₂O (19/1, 6.3 mL) was added DDQ (286 mg, 1.26 mmol) at 0°C, and the reaction mixture was stirred at r.t. for 2.5 hr. The reaction mixture was diluted with CH₂Cl₂ (30 mL), and the insoluble material was filtered off. Filtrate was concentrated under reduced pressure, and the residue was chromatographed on silica gel (AcOEt/hexane) to give 21 (416 mg, 91%) as a colorless oil. $[\alpha]_D^{20}$ +22.5° (c 1.12, CHCl₃): IR (neat) 3452, 3034, 2862, 1738, 1496, 1454cm⁻¹: ¹H-NMR (400MHz, CDCl₃) δ 2.18 (3H, s), 2.36 (1H, br, OH), 3.93 (1H, dd, J=3.7, 8.4Hz), 4.04 (1H, dd, J=1.6, 13.1Hz), 4.09 (1H, m), 4.12 (1H, dd, J=1.6, 8.4Hz), 4.31 (1H, dd, J=1.6, 13.1Hz), 4.69 (1H, d, J=11.3 Hz), 4.80 (1H, q, J=1.6Hz), 4.95 (1H, d, J=11.3Hz), 5.26 (1H, ddd, J=1.1, 1.2, 10.4Hz), 5.35 (1H, td, J=1.2, 17.3Hz), 5.63 (1H, s), 5.97 (1H, ddd, J=7.2, 10.4, 17.3Hz), 7.27-7.41 (8H, m), 7.52 (2H, m): Anal. Calcd for C₂₃H₂₆O₆: C, 69.33; H, 6.58%. Found: C, 69.09; H, 6.54%.

Phenyl 4,6-O-benzylidene-3-O-(4-methoxybenzyl)-1-thio-α-D-mannopyranoside (24): 7 (3.17 g, 8.80 mmol) was converted to 14, and 14 was reacted with CsF (1.36 g, 8.96 mmol) and MPMBr (1.85 g, 9.22 mmol) to give 24 (3.71 g, 88%) as white crystals. m.p. 100-101°C (Et₂O/hexane): $[\alpha]_D^{20}$ +209° (c 1.28, CHCl₃): IR (KBr) 3524, 2953, 2934, 2901, 2856, 1614, 1516, 1452, 1439cm⁻¹: ¹H-NMR (400MHz, CDCl₃) δ 2.82 (1H, d, J=1.3 Hz, OH), 3.82 (3H, s), 3.85 (1H, t, J=10.3Hz), 3.95 (H, dd, J=3.4, 9.5Hz), 4.15 (1H, t, J=9.5Hz), 4.20 (1H, dd, J=4.9, 10.3Hz), 4.25 (1H, ddd, J=1.2, 1.3, 3.4Hz), 4.33 (1H, ddd, J=4.9, 9.5, 10.3Hz), 4.68 (1H, d, J=11.4Hz), 4.82 (1H, d, J=11.4Hz), 5.59 (1H, d, J=1.2Hz), 5.62 (1H, s), 6.89 (2H, m), 7.26-7.54 (12H, m): Anal. Calcd for C₂₇H₂₈O₆S: C, 67.48; H, 5.87; S, 6.67%. Found: C, 67.49; H, 5.99; S, 6.68%.

Phenyl 3-O-(4-methoxybenzyl)-1-thio-α-D-mannopyranoside (25): 24 (4.37 g, 9.09 mmol) was dissolved in MeOH (40 mL) containing 1% conc. H₂SO₄, and the mixture was stirred at 0°C for 30 min.

MeOH (40 mL) saturated with NH₃ was added, and the mixture was concentrated under reduced pressure. The residue was chromatographed on silica gel (MeOH/CH₂Cl₂) to give **25** (3.22 g, 90%) as white crystals. $[\alpha]_D^{20}$ +164° (c 1.04, CHCl₃): IR (KBr) 3418, 2910, 1614, 1514cm⁻¹: ¹H-NMR (400MHz, CDCl₃/D₂O) δ 3.65 (1H, dd, J=3.0, 9.2Hz), 3.68 (1H, dd, J=2.0, 12.3Hz), 3.80 (3H, s), 3.81 (1H, m), 4.00-4.08 (2H, m), 4.11 (1H, dd, J=1.5, 3.0Hz), 4.54 (1H, d, J=11.2Hz), 4.62 (1H, d, J=11.2 Hz), 5.50 (1H, d, J=1.5Hz), 6.89 (2H, m), 7.25-7.43 (7H, m): *Anal*. Calcd for C₂₀H₂₄O₆S: C, 61.21; H, 6.16; S, 8.17%. Found: C, 61.18; H, 6.19; S, 8.07%.

Phenyl 2,4,6-tri-O-benzyl-3-O-(4-methoxybenzyl)-1-thio-α-D-mannopyranoside (26): 25 (3.04 g, 7.75 mmol) was benzylated in a manner similar to the preparation of 8 to give 26 (5.10 g, 99%) as a syrup. $[\alpha]_D^{20}$ +81.1° (c 1.21, CHCl₃): IR (neat) 3063, 3030, 2908, 2866, 1612, 1585, 1514, 1496, 1454cm⁻¹: ¹H-NMR (400MHz, CDCl₃) δ 3.75 (1H, dd, J=1.8, 10.9Hz), 3.81 (3H, s), 3.81-3.87 (2H, m), 3.96 (1H, dd, J=1.7, 3.0Hz), 4.04 (1H, t, J=9.5Hz), 4.27 (1H, ddd, J=1.8, 5.0, 9.5Hz), 4.48 (1H, d, J=12.0Hz), 4.51-4.57 (3H, m), 4.64 (2H, m), 4.72 (1H, d, J=12.3Hz), 4.90 (1H, d, J=10.8Hz), 7.64 (2H, m), 7.18-7.46 (18H, m): Anal. Calcd for C₄₁H₄₂O₆S: C, 74.29; H, 6.39; S, 4.84%. Found: C, 74.32; H, 6.60; S, 4.87%.

Phenyl 2,4,6-tri-*O*-benzyl-1-thio-α-D-mannopyranoside (27): 26 (5.10 g, 7.70 mmol) was reacted with DDQ (2.10 g, 9.24 mmol) in a manner similar to the preparation of 21 to give 27 (3.47 g, 83%) as syrup. [α] $_{\rm D}^{20}$ +117° (c 1.08, CHCl₃): IR (neat) 3431, 3063, 3030, 2916, 2866, 1454cm⁻¹: 1 H-NMR (400MHz, CDCl₃/D₂O) δ 3.74 (1H, dd, J=1.9, 10.9Hz), 3.77-3.88 (3H, m), 3.96-4.03 (2H, m), 4.29 (1H, ddd, J=1.9, 4.8, 9.7Hz),4.49 (1H, d, J=11.6Hz), 4.52 (1H, d, J=11.9Hz), 4.55 (1H, d, J=11.0Hz), 4.65 (1H, d, J=11.9Hz), 4.76 (1H, d, J=11.6Hz), 4.87 (1H, d, J=11.0Hz), 5.68 (1H, d, J=0.8Hz), 7.24-7.50 (20H, m).

Phenyl 2,4,6-tri-*O*-benzyl-3-*O*-carbamoyl-1-thio-α-D-mannopyranoside (28): Et₃N (2.88 mL, 20.7 mmol) was added to a mixture of 27 (4.48 g, 8.25 mmol) and 4-nitrophenyl chloroformate (3.83 g, 19.0 mmol) in THF (40 mL) at 0°C, and the mixture was stirred at r.t. for 6 hr. NH₃ gas was passed into the reaction mixture at 0°C for 1 hr, and the mixture was poured into sat. NaHCO₃. The products were extracted with AcOEt, and the organic layer was successively washed with 1*N* NaOH, brine, 1*N* HCl and brine, and dried over anhydrous Na₂SO₄, and concentrated under reduced pressure. The residue was chromatographed on silica gel (AcOEt/hexane) to give 28 (4.42 g, 92%) as a syrup. [α]_D²⁰ +79.5° (*c* 1.04, CHCl₃): IR (neat) 3493, 3381, 2868, 1730, 1585, 1454cm⁻¹: ¹H-NMR (400MHz, CDCl₃/D₂O) δ 3.72 (1H, dd, *J*=1.9, 11.0 Hz), 3.84 (1H, dd, *J*=4.6, 11.0Hz), 4.08 (1H, dd, *J*=9.2, 9.5Hz), 4.14 (1H, dd, *J*=2.1, 3.2Hz), 4.33 (1H, ddd, *J*=1.9, 4.6, 9.5Hz), 4.48 (1H, d, *J*=12.0Hz), 4.51 (1H, d, *J*=12.4Hz), 4.56 (1H, d, *J*=12.0Hz), 4.64-4.72 (5H, m), 5.13 (1H, dd, *J*=3.2, 9.2Hz), 5.58 (1H, d, *J*=2.1Hz), 7.21-7.49 (20H, m): *Anal.* Calcd for C₃₄H₃₅NO₆S: C, 69.72; H, 6.02; N, 2.39; S, 5.47%. Found: C, 69.50; H, 6.03; N, 2.44; S, 5.59%.

2,4,6-Tri-O-benzyl-3-O-carbamoyl-D-mannopyranose (29): **28** (4.26 g, 7.27 mmol) was reacted with NBS (2.59 g, 14.5 mmol) in a manner similar to the preparation of **18** to give **29** (2.67 g, 74%) as an anomeric mixture. SI-MS (3-NBA, KCl) m/z 533 ([M+1+K]+): Anal. Calcd for C₂₈H₃₁NO₇: C, 68.14; H, 6.33; N, 2.84%. Found: C, 67.87; H, 6.27; N, 2.73%.

2,4,6-Tri-O-benzyl-3-O-carbamoyl-α-D-mannopyranosyl trichloroacetimidate (30): 29 (2.57 g, 5.21 mmol) was reacted with CCl₃CN (0.61 mL, 6.10 mmol) and DBU (0.10 mL, 0.67 mmol) in CH₂Cl₂ (25 mL) at 0°C for 1 hr. Et₂O (30 mL) was added, and the organic layer was washed with brine, dried over anhydrous Na₂SO₄, and concentrated under reduced pressure. The residue was chromatographed on silica gel (AcOEt/hexane) to give 30 (3.07 g, 92%) as a syrup. IR (neat) 3493, 3381, 3339, 3032, 2918, 2870, 1728, 1672, 1599, 1496, 1454cm⁻¹: 1 H-NMR (400MHz, CDCl₃) δ 3.72 (1H, dd, 2 J=1.9, 11.3Hz), 3.82 (1H, dd, 2 J=4.0, 11.3Hz), 4.03 (1H, ddd, 2 J=1.9, 4.0, 9.7Hz), 4.12 (1H, dd, 2 J=2.4, 3.3Hz), 4.14 (1H, dd, 2 J=9.0, 9.7Hz), 4.52 (2H, m), 4.60-4.71 (5H, m), 4.76 (1H, d, 2 J=1.1Hz), 5.21 (1H, dd, 2 J=3.2, 9.0Hz), 6.37 (1H, d, 2 J=2.4Hz), 7.20-7.38 (20H, m), 8.59 (1H, s).

(1'R,2R,2'R,4R,5S)-5-Acetoxy-4-[1-benzyloxy-2-(2,4,6-tri-O-benzyl-3-O-carbamoyl- α -D-mannopyranosyl)oxy-3-butenyl]-2-phenyl-1,3-dioxane (31): BF3·OEt2 (0.61 mL, 5.0

mmol) was added to a mixture of 21 (988.0 mg, 2.48 mmol) and 30 (1.79 g, 2.81 mmol) in CH₂Cl₂ (20 mL) at -15°C, and the mixture was stirred at that temperature for 10 min. The reaction was quenched with sat. NaHCO₃ and the product was extracted with AcOEt. The organic layer was washed with brine, dried over anhydrous Na₂SO₄, and concentrated under reduced pressure. The residue was chromatographed on silica gel (AcOEt/hexane) to give 31 (1.78 g, 82%) as a syrup. $[\alpha]_D^{20}$ +33.9° (c 1.04, CHCl₃): IR (neat) 3493, 3373, 3032, 2914, 2868, 1736, 1599, 1496, 1454cm⁻¹: 1 H-NMR (400MHz, CDCl₃) δ 2.10 (3H, s), 3.62 (1H, br, J=10.8Hz), 3.76 (1H, dd, J=3.7, 10.8Hz), 3.80 (1H, ddd, J=1.5, 3.7, 9.5Hz), 3.88 (1H, dd, J=2.0, 3.2 Hz), 3.91 (1H, dd, J=1.9, 8.3Hz), 4.01 (1H, dd, J=1.5, 13.1Hz), 4.06 (2H, m), 4.20 (1H, br, J=7.2Hz), 4.30 (1H, dd, J=1.5, 13.1Hz), 4.45-4.73 (11H, m), 5.04 (1H, d, J=2.0Hz), 5.16 (1H, dd, J=3.2, 9.6Hz), 5.18 (1H, br, J=10.4Hz), 5.25 (1H, br, J=17.4Hz), 5.59 (1H, s), 5.93 (1H, ddd, J=7.2, 10.4, 17.4Hz), 7.19-7.41 (25H, m): Anal. Calcd for C₅₁H₅₅NO₁₂: C, 70.09; H, 6.34; N, 1.60%. Found: C, 69.84; H, 6.54; N, 1.49%.

(1'R,2R,2'R,4R,5S)-4-[1-benzyloxy-2-(2,4,6-tri-*O*-benzyl-3-*O*-carbamoyl-α-D-mannopyranosyl)oxy-3-butenyl]-5-hydroxy-2-phenyl-1,3-dioxane (32): A mixture of 31 (148.4 mg, 0.170 mmol) and K₂CO₃ (44.0 mg) in MeOH (2 mL) was stirred at 60°C for 10 min. After being neutralized with AcOH, the mixture was concentrated under reduced pressure. The residue was chromatographed on silica gel (AcOEt/hexane) to give 32 (131.4 mg, 93%) as a syrup. $[\alpha]_D^{20}$ +12.4° (*c* 1.04, CHCl₃): IR (neat) 3398, 2922, 2851, 1718, 1655, 1560, 1496, 1458cm⁻¹: ¹H-NMR (400MHz, CDCl₃) δ 3.02 (1H, d, J=10.0Hz, OH), 3.58 (1H, br, J=10.0Hz), 3.64 (1H, dd, J=1.9, 10.9Hz), 3.77 (1H, dd, J=4.3, 10.9Hz), 3.85 (1H, dd, J=3.0, 7.4Hz), 3.88 (1H, dd, J=2.1, 3.2Hz), 3.89 (1H, m), 3.94 (1H, dd, J=1.0, 7.4Hz), 3.95 (1H, dd, J=1.2, 12.0Hz), 4.03 (1H, t, J=9.5Hz), 4.11 (1H, dd, J=1.9, 12.0Hz), 4.39 (1H, dd, J=3.0, 7.5Hz), 4.46-4.59 (6H, m), 4.62-4.73 (4H, m), 5.10 (1H, d, J=2.1Hz), 5.20 (1H, dd, J=3.2, 9.2 Hz), 5.22 (1H, br, J=10.4Hz), 5.29 (1H, br, J=17.2Hz), 5.55 (1H, s), 6.04 (1H, ddd, J=7.5, 10.4, 17.2 Hz), 7.20-7.53 (25H, m).

3-*O*-Benzyl-4,6-*O*-benzylidene-2-*O*-(2,4,6-tri-*O*-benzyl-3-*O*-carbamoyl-α-D-manno-pyranosyl)-L-gulopyranose (33): 32 (1.61 g, 1.94 mmol) was reacted with O₃ in a manner similar to the preparation of 13 to give 33 (1.51 g, 94%) as an anomeric mixture ($\alpha/\beta=5/3$). SI-MS (3-NBA, NaCl) m/z 856 ([M+Na]+); *Anal.* Calcd for C₄₈H₅₁NO₁₂: C, 69.13; H, 6.16; N, 1.68%. Found: C, 68.83; H, 6.28; N, 1.55%.

1,3,4,6-Tetra-O-acetyl-2-O-(2,4,6-tri-O-acetyl-3-O-carbamoyl- α -D-mannopyranosyl)-L-gulopyranose (34): 33 (158.4 mg, 0.190 mmol) in MeOH (3 mL) was stirred under H₂ in the presence of 20% Pd(OH)₂ (15.4 mg) to give unprotected mannogulose (69.1 mg, 94%) as a solid. Mannogulose (266.9 mg, 0.692 mmol) prepared in a similar manner was dissolved in pyridine (5 mL), and was added Ac₂O (0.80 mL, 8.5 mmol). The mixture was stirred at r.t. for 10 hr, and was poured into sat. NaHCO₃. The product was extracted with Et₂O. The organic layer was washed with brine, dried over anhydrous Na₂SO₄, and concentrated under reduced pressure. The residue was chromatographed on silica gel (AcOEt/hexane) to give 34 (446.9 mg, 95%) as a syrup. $[\alpha]_D^{20} + 35.5^{\circ}$ (c 0.95, CHCl₃): IR (neat) 3481, 3381, 2959, 1749, 1608cm⁻¹: ¹H-NMR (400MHz, CDCl₃) & 2.05 (3H, s), 2.06 (3H, s), 2.12 (3H, s), 2.13 (3H, s), 2.14 (3H, s), 2.15 (3H, s), 2.20 (3H, s), 4.00 (1H, dd, J=3.3, 8.4Hz), 4.06-4.29 (5H, m), 4.36 (1H, m), 4.64 (2H, br), 5.00 (1H, d, J=1.6Hz), 5.02 (1H, dd, J=1.4, 3.9Hz), 5.08 (1H, dd, J=3.4, 10.0Hz), 5.12 (1H, dd, J=1.8, 3.4Hz), 5.26 (1H, t, J=10.0Hz), 5.44 (1H, t, J=3.4Hz), 5.89 (1H, d, J=8.4Hz).

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